ІТЕМ ТО СНЕСК	IMPORTANCE	CHECKLIST
EXPERIMENTAL DESIGN Definition of experimental and control groups	E	
Number within each group	E D	
Assay carried out by core lab or investigator's lab? Acknowledgement of authors' contributions	D	
SAMPLE Description	E	
Volume/mass of sample processed	D	
Microdissection or macrodissection Processing procedure	E E	
If frozen - how and how quickly?	E	
If fixed - with what, how quickly? Sample storage conditions and duration (especially for FFPE samples)	E E	
NUCLEIC ACID EXTRACTION Procedure and/or instrumentation	E	
Name of kit and details of any modifications	E	
Source of additional reagents used Details of DNase or RNAse treatment	D E	
Contamination assessment (DNA or RNA)	E	
Nucleic acid quantification Instrument and method	E E	
Purity (A260/A280)	D	
Yield RNA integrity method/instrument	D E	
RIN/RQI or Cq of 3¹ and 5¹ transcripts Electrophoresis traces	E D	
Inhibition testing (Cq dilutions, spike or other)	E E	
REVERSE TRANSCRIPTION Complete reaction conditions	E	
Amount of RNA and reaction volume	E	
Priming oligonucleotide (if using GSP) and concentration Reverse transcriptase and concentration	E E	
Temperature and time	E	
Manufacturer of reagents and catalogue numbers Cqs with and without RT	D D*	
Storage conditions of cDNA	D	
qPCR TARGET INFORMATION If multiplex, efficiency and LOD of each assay.	E	
Sequence accession number Location of amplicon	E D	
Amplicon length	E	
In silico specificity screen (BLAST, etc) Pseudogenes, retropseudogenes or other homologs?	E D	
Sequence alignment	D	
Secondary structure analysis of amplicon Location of each primer by exon or intron (if applicable)	D E	
What splice variants are targeted?	E	
What splice variants are targeted? qPCR OLIGONUCLEOTIDES Primer sequences		
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Table 1. MIQE checklist for authors, reviewers and editors. All essential information (E) must be submitted with the manuscript. Desirable information (D) should be submitted if available. If using primers obtained from RTPrimerDB, information on qPCR target, oligonucleotides, protocols and validation is available from that source.

^{*:} Assessing the absence of DNA using a no RT assay is essential when first extracting RNA. Once the sample has been validated as RDNA-free, inclusion of a no-RT control is desirable, but no longer essential.

^{**:} Disclosure of the probe sequence is highly desirable and strongly encouraged. However, since not all commercial pre-designed assay vendors provide this information, it cannot be an essential requirement. Use of such assays is advised against.